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Suppression of reproductive characteristics of the invasive plant Mikania micrantha by sweet potato competition

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Abstract

Background: As a means of biologically controlling *Mikania micrantha* H.B.K. in Yunnan, China, the influence of sweet potato [*Ipomoea batatas* (L.) Lam.] on its reproductive characteristics was studied. The trial utilized a de Wit replacement series incorporating six ratios of sweet potato and *M. micrantha* plants in 25 m² plots over 2 years.

Results: Budding of *M. micrantha* occurred at the end of September; flowering and fruiting occurred from October to February. Flowering phenology of *M. micrantha* was delayed (P < 0.05), duration of flowering and fruiting was reduced (P < 0.05) and duration of bud formation was increased (P < 0.05) with increasing proportions of sweet potato. Reproductive allocation, reproductive investment and reproductive index of *M. micrantha* were significantly reduced (P < 0.05) with increasing sweet potato densities. Apidae bees, and Calliphoridae or Syrphidae flies were the most abundant visitors to *M. micrantha* flowers. Overall flower visits decreased (P < 0.05) as sweet potato increased. Thus the mechanism by which sweet potato suppressed sexual reproduction in *M. micrantha* was essentially two-fold: causing a delay in flowering phenology and reducing pollinator visits. The number, biomass, length, set rate, germination rate, and 1000-grain dry weight of *M. micrantha* seeds were suppressed (P < 0.05) by sweet potato competition. With proportional increases in sweet potato, sexual and asexual seedling populations of *M. micrantha* were significantly reduced (P < 0.05). The mortality of both seedling types increased (P < 0.05) with proportional increases in sweet potato.

Conclusions: These results suggest that sweet potato significantly suppresses the reproductive ability of the invasive species *M. micrantha*, and is a promising alternative to traditional biological control and other methods of control. Planting sweet potato in conjunction with other control methods could provide a comprehensive strategy for managing *M. micrantha*. The scenario of controlling *M. micrantha* by utilizing a crop with a similar growth form may provide a useful model for similar management strategies in other systems.

Keywords: Biological control, *Mikania micrantha*, Sweet potato, Competition, Flowering and fruiting phenology, Reproductive suppression

Background

Plant invasions have received attention on a global scale. Invasive plants have caused great economic harm,

biodiversity loss, environmental problems, and even human and animal health issues [1, 2]. Currently, methods to control invasive plants have been widely investigated and practiced, including mechanical [3–5], chemical [6, 7] and biological control [8–11]. However, mechanical measures may potentially accelerate invasions [3], chemical measures can be detrimental to nontarget species and environment health [12–14], and traditional biological control measures via introduction

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of pathogens, parasites, and predators against invaders are expensive and pose risks to ecosystem integrity [15, 16]. Thus, as a potential alternative to traditional biological control which generally employs insects or pathogens, replacement control relies on growth characteristics of one or more plants to suppress exotic plants, simultaneously reducing damage caused by the invasive species and improving local natural ecosystem health by reducing the potential for invasive plants to spread beyond agricultural fields. Adoption of this alternative method has received considerable attention in recent years [17–21].

Mikania micrantha H.B.K. is a rapidly-growing perennial creeping vine belonging to the family Asteraceae native to Central and South America [22]. The vine has been listed among the top 100 worst invasive species [22, 23] and as one of the top 10 worst weeds in the world [22]. M. micrantha is present in tropical Asia, parts of Papua New Guinea, Indian Ocean islands, Pacific Ocean Islands, and Florida in the US [22, 24, 25]. It has colonized a broad range of farming systems and forest lands, banks of streams and rivers, roadsides and railway tracks, pastures, and open disturbed areas [22], leading to serious economic loss, biodiversity loss and negative environmental impacts [7, 18, 22, 26, 27].

To explore ecological methods for managing M. micrantha, biological control measures through replacement control with high value species (e.g., local food, native species and/or cash crops) have been investigated [18, 28-31]. In 2006 and 2007, sweet potato [Ipomoea batatas (L.) Lam.: Convolvulaceae], an important locally grown cash crop native to the American tropics, was observed to inhibit M. micrantha growth in Longchuan County of Yunnan Province, China in sweet potato fields where M. micrantha occurred [30]. Subsequent studies examined the effects of a local crop, sweet potato, on M. micrantha growth and soil nutrients [18]. Sweet potato exhibited greater competitive ability than M. micrantha, with plant height, branch, leaf, stem node, adventitious root, and biomass of M. micrantha suppressed significantly; furthermore sweet potato also demonstrated higher levels of nutrient uptake than M. micrantha. Moreover, flowering of M. micrantha was significantly suppressed in mixed culture with sweet potato and with decreasing proportions of M. micrantha, the competitiveness of sweet potato increased at a rate exceeding what would be predicted by the increase in relative density [18]. However, no literature is available on the effects of sweet potato competition on the entire suite of reproductive characteristics of *M. micrantha*.

Building on our former studies [18, 30, 31], the present research examined how sweet potato suppressed reproductive characteristics of *M. micrantha* in Yunnan Province, China. Previous reports did not refer to impacts on

reproductive characteristics, so this is the first report of how sweet potato competition affects characteristics such as flowering and seed production in *M. micrantha*. These findings are important to further elucidate the competitive interaction and mechanisms between sweet potato and *M. micrantha* and provide insights for similar ecological control methods that could be applied to other invasive alien species.

Methods

Study site

The study site was located in Longchuan County $(24^{\circ}08'-24^{\circ}39' \text{ N}, 97^{\circ}17'-97^{\circ}39' \text{ E})$, in the western end of Yunnan Province, Southwest China. This area is characterized by a typical tropical climate, having a rainy season featuring heavy rainfall with 90 % relative humidity alternating with a dry season [30]. Rainfall averages 15450 mm per year and the annual mean temperature is 18.9 °C. Recently, the range of *M. micrantha* has been expanding rapidly within Longchuan County, as the plant has invaded agricultural areas and forest margins [7].

Study species

Mikania micrantha is one of the most serious invasive species in Longchuan County where this study took place. This perennial weed exhibits a climbing growth form in forests, orchards and shrublands, but on roadsides, in open wasteland areas without crops, and other areas without woody vegetation, it takes on a prostrate form. It has infested sugarcane, orange, banana, coffee, pineapple, bamboo, sweet potato, maize crops, as well as artificial pasture and secondary forest in the study area [7]. M. micrantha can invade disturbed environments via light weight wind-dispersed seeds that are produced in great numbers, as high as 170,000 m⁻² [32]. At a local level, vegetative reproduction is responsible for most population growth as facilitated by rooting of stem fragments [3].

Sweet potato, native to the American tropics, is one of the main food and cash crops in tropical and subtropical regions of Yunnan Province. It is also grown in many other regions of China and other subtropical or warm-temperate regions of the world as a food source. In Longchuan County, local villagers have grown it for over 100 years [30]. This herbaceous perennial vine usually exhibits a prostrate growth form in agricultural areas, so its niche is similar to that of *M. micrantha*. Because of its purple root, it is also known as purple sweet potato. The aboveground parts of the plant are used for livestock fodder and its roots are used for human eating. It is propagated by seed or by clonal means, with 20–50 cm fragments with 3–5 nodes typically planted [33].

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Experiment design and data collection

The experiments were conducted during the May, 2014-October, 2015 growing season within maize and sweet potato intercropping land in the vicinity of Zhangfeng Town, Longchuan County, utilizing a de Wit replacement series method [34]. On 7 May 2014, whole M. micrantha plants (including roots) were collected from a M. micrantha population located in a nearby forest margin and whole sweet potato plants were collected from farmland, respectively. To ensure relative uniformity among the experimental stock, one-node segments (fresh weight 3.0–3.5 g, 7–8 cm pieces) were taken from central stem portions of relatively young plants of similar size from both species. The segments were placed in Hoagland's solution [35] and grown for 10 days. On 17 May 2014, the sprouts derived from cuttings of both species were transplanted in the field test plots. Six ratios of sweet potato and M. micrantha plants were utilized (3:1, 2:1, 1:1, 1:2, 1:3, 0:4) while maintaining a constant planting density of 20 plants m⁻² (0.25 \times 0.20 m spacing). All plots were arranged in a complete randomized design with 4 replicates utilizing 25 m² plots (5 \times 5 m). All sweet potato and M. micrantha plants were distributed evenly within the plot. During the experiment, the two species exhibited prostrate growth. The plots were not weeded and no fertilizers were used.

From October 2014 to February 2015, flowering and fruiting phenology were recorded at 3 days intervals for M. micrantha, including the dates of initial and last budding, flowering, fruiting and seed set. During peak flowering times, we marked 20 inflorescences of M. micrantha in each plot and recorded the number of flower visits per insect visitor on each inflorescence between 9:00 and 18:00 (time of maximum pollinator activity) for two continuous days with all plots monitored simultaneously over the same two days. According to [36] each flower visitor was classified as belonging to Apidae, Calliphoridae, Syrphidae (generally the most frequent visitors to M. micrantha), or another pollinator group beyond these three taxa. At the same time, twenty plants of each species were selected randomly and harvested within the middle region of each plot. M. micrantha plants were carefully removed and separated, and number of inflorescences and flowers were recorded. The fresh weight of inflorescences and flowers and total biomass of each M. micrantha plant were measured. On 28 January 2015, seed production of M. micrantha was measured in the study plots after flowering had waned, but prior to seed dispersal. Another twenty plants of each species were selected randomly and harvested within the middle region of each plot. The number, size (length) and dry weight of M. micrantha seeds were measured. Sixty days later, germination rates of *M. micrantha* seeds from each plot were tested in the laboratory.

During the spring, summer and fall of 2015 (March–October), four small quadrats $(1 \times 1 \text{ m})$ were selected randomly and marked in each plot. Seedlings were identified as either produced from germinating seeds (sexual) or vegetative growth (asexual). The number of new sexual and asexual M. micrantha seedlings was monitored in each quadrat monthly. We did not remove the seedlings that were counted but rather kept track of the total that emerged through the season, month by month. Seedling mortality was also recorded for the M. micrantha that emerged.

Data analyses

Reproductive characteristics of M. micrantha [37, 38] were calculated in each plot with the following parameters: (1) Reproductive allocation $(g \cdot g^{-1}) = inflorescence$ biomass/total biomass of each plant, (2) Reproductive investment $(g \cdot g^{-1}) = flower$ biomass/total biomass of each plant, (3) Reproductive index $(g \cdot g^{-1}) = flower$ biomass/inflorescence biomass of each plant, (4) Reproductive ratio (flower·mg⁻¹) = flower number/inflorescence biomass of each plant and (5) Reproductive efficiency index (flower·mg⁻¹) = flower number/total biomass of each plant.

All growth variables (flowering, bud formation and fruiting duration, inflorescence number, flower number, germination, and biomass of inflorescences, and flowers and seeds) of *M. micrantha* plants were analyzed by analysis of variance (one-way ANOVA) using IBM SPSS 22.0 software. If significant differences were detected with the ANOVA, Tukey's HSD, Post Hoc Multiple Comparisons, Homogeneity of Variance tests were used to detect differences among treatments at a 5 % level of significance.

Results

Reproductive phenology of Mikania micrantha

Budding of M. micrantha occurred at the end of September, and flowering and fruiting occurred from October to February in our study area (Table 1). The peak bloom occurred between mid-November and early December. Fruiting started as early as the end of November, and almost all fruits had dropped by early February. The flowers opened throughout the day, but the majority did so in the morning. In monoculture, the duration of bud formation, flowers and fruits of M. micrantha was 27.25 ± 0.05 d, 79.75 ± 1.71 d, and 75.75 ± 1.71 d, respectively (Table 1). With increased sweet potato: M. micrantha ratios, the initial date of budding, flowering and fruiting of M. micrantha was significantly delayed, and the duration of flowering and fruiting of

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Table 1 Flowering and fruiting phenology of *Mikania micrantha* growing as a monoculture or under mixed culture conditions

Variables	Ratios (sweet potato: M. micrantha)							
	3:1	2:1	1:1	1:2	1:3	0:4		
Initial budding date	10 October	5 October	2 October	28 September	26 September	26 September		
Initial flowering date	16 November	12 November	4 November	27 October	24 October	24 October		
Initial fruiting date	20 November	16 November	11 November	4 November	2 November	1 November		
Duration of bud formation (d)	$37.75 \pm 0.96a$	$34.75 \pm 0.96b$	$31.25 \pm 0.96c$	$30.25 \pm 0.96c$	$28.25 \pm 0.96 \text{cd}$	$27.25 \pm 0.05 d$		
Duration of flowering (d)	$48.25 \pm 1.26f$	$54.75 \pm 1.50e$	$60.50 \pm 1.29 d$	$68.25 \pm 2.22c$	$73.75 \pm 2.06b$	79.75 ± 1.71a		
Duration of fruiting (d)	$48.25 \pm 1.89e$	$55.00 \pm 2.16d$	$59.50 \pm 1.29c$	66.50 ± 1.29 b	70.00 ± 1.63 b	75.75 ± 1.71a		

Data are expressed as mean \pm standard deviation. The different letters within same row signify significantly different at P < 0.05

M. micrantha significantly declined, but duration of bud formation of *M. micrantha* was markedly increased (P < 0.05).

A total of about 47 flower visits of M. micrantha were observed for each inflorescence per day, with Apidae (bees), Calliphoridae (flies), Syrphidae (flies) and other pollinators accounting for 61.2, 22.0, 12.4, and 4.4 % of total visits, respectively, in monoculture (Table 2). Overall flower visiting behavior and visitation rate of Apidae, Calliphoridae and Syrphidae per inflorescence were substantially reduced with increasing proportions of sweet potato (P < 0.05).

Reproductive characteristics of Mikania micrantha

In monoculture, the biomass of plant, inflorescences and flowers of M. micrantha was 301.61 ± 5.19 g, 54.66 ± 1.14 g, and 40.99 ± 1.01 g, respectively; the inflorescence and flower numbers of M. micrantha were 2647.8 ± 55.3 and $10,587.7 \pm 239.3$, respectively. In mixed culture, the total biomass of plant, biomass of inflorescences and flowers, and numbers of inflorescences and flowers of M. micrantha were significantly (P < 0.05) suppressed with decreasing proportions of M. micrantha (Table 3). With proportional increases in sweet potato, reproductive allocation, reproductive

investment and reproductive index of M. micrantha were significantly lower (P < 0.05). The reproductive ratio of M. micrantha did not differ significantly among treatments (Table 3). With decreasing proportions of M. micrantha, the reproductive efficiency index of M. micrantha was reduced to a certain extent by sweet potato but the trend was not clear.

For a ratio of sweet potato to M. micrantha of 3:1, the number and biomass of M. micrantha seeds were reduced by a factor of more than 100 compared to M. micrantha in monoculture, i.e., $17,632.6 \pm 479.8$ vs. 171.7 ± 4.3 and 1.772 ± 0.042 g vs. 0.014 ± 0.000 g, respectively (Table 4). The number, biomass, length, set rate, germination rate, and 1000-grain dry weight of M. micrantha seeds were significantly suppressed (P < 0.05) with decreasing proportions of M. micrantha.

Seedling population dynamics of Mikania micrantha

Sexual seedling populations of *M. micrantha* germinated for 6 months (March–August), primarily occurring between May–June. Asexual seedling populations first arose in March, and then increased in density monthly from March to October. In monoculture, sexual population and asexual population densities were 89.25 \pm 4.35 m⁻² and 134.75 \pm 4.99 m⁻², respectively

Table 2 Total number of visits (visits per day and inflorescence) by the four pollinator groups to *Mikania micrantha* growing as a monoculture or under mixed culture conditions

Pollinators	Ratios (sweet potato: <i>M. micrantha</i>)								
	3:1	2:1	1:1	1:2	1:3	0:4			
Apidae	8.75 ± 0.95d	10.50 ± 1.11d	13.50 ± 1.04c	24.50 ± 1.14b	27.50 ± 1.43a	28.50 ± 1.40a			
Calliphoridae	$4.92 \pm 0.70c$	$6.00 \pm 0.63c$	6.50 ± 0.50 bc	$8.02 \pm 0.75b$	$9.74 \pm 0.88a$	$10.26 \pm 0.96a$			
Syrphidae	$1.26 \pm 0.46c$	$1.54 \pm 0.45c$	$2.28 \pm 0.40c$	$3.76 \pm 0.39b$	$5.63 \pm 0.54a$	$5.75 \pm 0.85a$			
Other pollinator	$2.53 \pm 0.48a$	$1.63 \pm 0.57a$	$1.76 \pm 0.41a$	$2.23 \pm 0.73a$	$1.77 \pm 0.92a$	$2.04 \pm 0.67a$			
Total	$17.46 \pm 1.19d$	19.66 ± 2.60 d	$24.04 \pm 1.76c$	38.50 ± 1.55 b	$44.63 \pm 2.13a$	$46.55 \pm 0.95a$			

 $Data\ are\ expressed\ as\ mean\ \pm\ standard\ deviation.\ Different\ letters\ within\ the\ same\ row\ signify\ significant\ differences\ at\ P<0.05$

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Table 3 Flowering characteristics of Mikania micrantha growing as a monoculture or under mixed culture conditions

Variables	Ratios (sweet potato: M. micrantha)							
	3:1	2:1	1:1	1:2	1:3	0:4		
Total biomass (g)	25.16 ± 1.10f	35.88 ± 0.96e	54.80 ± 0.80d	82.21 ± 1.36c	104.41 ± 2.62b	301.61 ± 5.19a		
Flower number	$429.9 \pm 10.3 f$	1168.3 ± 82.7e	$2486.2 \pm 62.9 d$	$3508.8 \pm 53.8c$	$4705.9 \pm 106.5b$	10,587.7 ± 239.3a		
Inflorescence number	$108.3 \pm 3.7 f$	289.8 ± 14.1e	$613.9 \pm 12.1d$	868.8 ± 16.1c	$1179.4 \pm 22.5b$	$2647.8 \pm 55.3a$		
Flower biomass (g)	$0.63 \pm 0.03e$	$1.55 \pm 0.04e$	$3.86 \pm 0.06d$	$7.32 \pm 0.06c$	$11.39 \pm 0.29b$	$40.99 \pm 1.01a$		
Inflorescence biomass (g)	$1.24 \pm 0.04e$	$2.28 \pm 0.07e$	$6.86 \pm 0.09 d$	$11.21 \pm 0.13c$	$14.95 \pm 0.27b$	$54.66 \pm 1.14a$		
Reproductive allocation (g·g ⁻¹)	$0.049 \pm 0.004 f$	$0.064 \pm 0.002e$	$0.125 \pm 0.002d$	$0.136 \pm 0.001c$	$0.143 \pm 0.002b$	$0.181 \pm 0.002a$		
Reproductive investment $(g \cdot g^{-1})$	$0.025 \pm 0.002f$	$0.043 \pm 0.001e$	$0.071 \pm 0.001 d$	$0.089 \pm 0.002c$	0.109 ± 0.001 b	$0.136 \pm 0.002a$		
Reproductive index (g·g ⁻¹)	$0.512 \pm 0.012d$	0.678 ± 0.005 b	$0.563 \pm 0.003c$	0.653 ± 0.010 b	$0.762 \pm 0.012a$	$0.750 \pm 0.018a$		
Reproductive ratio (flower⋅mg ⁻¹)	$3.970 \pm 0.081a$	$4.028 \pm 0.089a$	$4.049 \pm 0.025a$	$4.039 \pm 0.025a$	$3.990 \pm 0.015a$	$3.999 \pm 0.041a$		
Reproductive efficiency index (flower \cdot mg ⁻¹)	$0.017 \pm 0.001d$	$0.033 \pm 0.002c$	$0.045 \pm 0.001a$	0.043 ± 0.000 b	$0.045 \pm 0.001 \mathrm{ab}$	$0.035 \pm 0.000c$		

Data are expressed as mean \pm standard deviation. The different letters within same row signify significant differences at P < 0.05

Table 4 Characteristics of Mikania micrantha seed growing as a monoculture or under mixed culture conditions

Variables	Ratios (sweet potato: M. micrantha)							
	3:1	2:1	1:1	1:2	1:3	0:4		
Seed number	171.7 ± 4.3f	702.9 ± 8.2e	1995.1 ± 54.9d	4066.9 ± 94.8c	6797.6 ± 92.2b	17,632.6 ± 479.8a		
Seed biomass (g)	$0.014 \pm 0.000f$	$0.058 \pm 0.001e$	$0.171 \pm 0.004d$	$0.362 \pm 0.008c$	$0.654 \pm 0.011b$	$1.772 \pm 0.042a$		
Seed length (mm)	$0.750 \pm 0.008f$	$0.808 \pm 0.013e$	$0.860 \pm 0.008d$	$0.900 \pm 0.012c$	1.058 ± 0.013 b	$1.290 \pm 0.014a$		
Seed set rate (%)	$7.99 \pm 0.26 f$	$12.07 \pm 0.76e$	$16.05 \pm 0.13d$	$23.18 \pm 0.27c$	$28.90 \pm 0.53b$	$33.31 \pm 0.64a$		
Germination rate (%)	$16.88 \pm 0.48 f$	$20.63 \pm 0.63e$	$25.13 \pm 0.85d$	$32.63 \pm 1.65c$	$53.13 \pm 1.60b$	$68.25 \pm 1.04a$		
1000-grain dry weight (g)	$0.078 \pm 0.001f$	$0.082 \pm 0.001e$	$0.086 \pm 0.001d$	$0.089 \pm 0.001c$	0.096 ± 0.001 b	0.101 ± 0.001a		

Data are expressed as mean \pm standard deviation. Different letters within the same row signify significant differences at P < 0.05

Discussion

Our research showed that the biomass, flowering, seed and seedling characteristics of *M. micrantha* were reduced when grown in association with sweet potato. A previous study showed that plant height, branch, leaf, stem node, adventitious root, and biomass of *M. micrantha* were suppressed significantly by sweet potato competition [18]. The present study found that the biomass of plant, inflorescences, flowers, and seeds of *M. micrantha*

were also significantly suppressed with increasing proportions of sweet potato. Moreover, with decreasing number and biomass of inflorescences and flowers, the reproductive allocation, reproductive investment and reproductive index of *M. micrantha* were also significantly reduced. The net result of the presence of sweet potato was reduced reproductive potential of *M. micrantha* and like other invasive species, its reproductive ability, including flowering characteristics, seed dispersal and seed germination parameters, is associated with its invasiveness [39–43].

Flowering phenology is affected by number, timing and duration of flowers [44]. These factors are not only constrained by genetics and phylogeny, but also affected by environmental conditions, such as sunlight, temperature, nutrients, and competition [42]. The present study found that flowering phenology of *M. micrantha* was significantly delayed, duration of flowering and fruiting was significantly reduced and duration of bud formation was markedly increased with increased sweet potato proportions.

Along with reduced biomass and delayed flowering, another major factor reducing reproductive output in *M*.

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Table 5 Population densities m⁻² from sexual and asexual reproduction of *Mikania micrantha* growing as a monoculture or under mixed culture conditions

Variables	Ratios (sweet potato: M. micrantha)							
	3:1	2:1	1:1	1:2	1:3	0:4		
Total population	42.00 ± 2.83f	65.25 ± 2.99e	102.50 ± 3.70d	166.75 ± 6.02c	191.50 ± 4.51b	224.00 ± 6.06a		
Sexual population	$9.25 \pm 1.71f$	$21.00 \pm 2.16e$	$31.50 \pm 2.08d$	$55.75 \pm 3.30c$	$70.50 \pm 1.29b$	$89.25 \pm 4.35a$		
Asexual population	$32.75 \pm 1.71f$	$44.25 \pm 2.63e$	$71.00 \pm 2.83d$	$111.00 \pm 3.92c$	$121.00 \pm 3.37b$	$134.75 \pm 4.99a$		
Sexual mortality rate (%)	$0.629 \pm 0.092a$	$0.596 \pm 0.080a$	$0.438 \pm 0.036b$	0.340 ± 0.035 bc	$0.291 \pm 0.022c$	$0.241 \pm 0.019c$		
Asexual mortality rate (%)	$0.358 \pm 0.034a$	$0.265 \pm 0.008b$	$0.152 \pm 0.019c$	$0.077 \pm 0.014d$	0.056 ± 0.008 de	$0.030 \pm 0.001e$		

Data are expressed as mean + standard deviation. The different letters within same row signify significant differences at P < 0.05

micrantha observed in our study was the negative impact of sweet potato on pollinator visitation. M. micrantha depends on insects for sexual reproduction as it is selfincompatible [45]. For insect-pollinated plant species, competition for pollinators may lead to changes in visitation frequency or pollinator composition [46-48] and consequently, a lowered reproductive output [48]. In this study, species of Apidae (bee species), Calliphoridae (flies), and Syrphidae (flies) were the most abundant visitors to M. micrantha flowers and were observed to have the longest foraging time of all floral visitors, which is consistent with the results of other studies [36, 49]. Overall flower visits of Apidae, Calliphoridae and Syrphidae per inflorescence were reduced significantly with increasing proportions of sweet potato. From October to February in the study area, the temperature gradually became lower and the delayed flowering phenology of M. micrantha corresponded with reduced insect activity. Moreover, in mixed culture, 70–90 % of M. micrantha stems and leaves were covered by sweet potato [18], thus reducing insect visitation via diminished visibility of M. micrantha flowers. Because flowering in both sweet potato and M. micrantha occurs at virtually the same time, pollinators visited both species during the monitoring period; however, because the number of flowers per shoot was at least 15 times greater for M. micrantha than sweet potato [18], the main influence on pollinator visitation was M. micrantha flower number. M. micrantha can produce a large number of flowers and small, light, and wind-dispersed seeds [32]. The negative correlation we observed between seed set and pollinator visitation in *M*. micrantha is consistent with the commonly observed link between pollinator visitation rate and seed set [50].

Nutrient availability also influences reproductive output by *M. micrantha*, with fewer flowers, lower seed setting percentage, lower 1000-grain weight and shorter flowering duration observed in plants growing in nutrient-deficient soils with suboptimal fertility, but soils with an overabundance of nutrients (e.g., silt from ponds or

dump sites) also resulted in fewer flowers and low seed set [51]. Meanwhile, plants growing in an open habitat had more flowers with longer flowering duration, and under shade the 1000-grain weight was shown to have a slight increase, but light that was too bright or too dim was not conducive to seed set [51, 52]. Light was found to affect fruiting; for example, a photoperiod of 12 h/day resulted in 68.4 % of flowers producing fruit [53]. The present study found that the number, biomass, length, set rate, germination rate, and 1000-grain dry weight of M. micrantha seeds was significantly suppressed with decreasing proportions of *M. micrantha*. This is because M. micrantha plants covered by dense carpets of sweet potato received fewer pollinator visits and produced fewer seeds. Furthermore, sweet potato exhibited greater absorption of soil nutrients than M. micrantha [18]; the resulting lack of nutrients likely lead to reduced 1000grain weight and germination rate of *M. micrantha* in the presence of sweet potato.

The potential for high levels of sexual and/or vegetative reproduction by M. micrantha is formidable [22]. It has transient soil seed bank and persistent soil seed bank, and some seeds would germinate given ideal germination conditions such as season, temperature, moisture; otherwise the seeds would remain dormant [54, 55]. Large numbers of seeds of M. micrantha were concentrated primarily in the 0-5 cm soil layer, which contained 98 % of the total seeds present in the soil [55]. Vegetative propagation of M. micrantha from stem fragments that root easily at the nodes and from vegetative ramets arising from rosettes can be considered at least as important as reproduction by seeds [22, 56]. The seedling is the most vulnerable stage in the life history of *M. micrantha* and seedlings suffer a high level of mortality under natural conditions [54]. The present study found that with proportional increases in sweet potato, both sexual and asexual seedling populations of M. micrantha were significantly suppressed, corresponding to increased mortality with increasing levels of sweet potato competition.

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Thus, the best time to control *M. micrantha* is during the seedling period and control measures should be comprehensive involving both herbicides and appropriate cultural techniques [31].

Conclusion

The competitive advantage of sweet potato over M. micrantha could be used to reduce M. micrantha growth and reproductive ability in tropical and subtropical agricultural regions suitable for cultivation of sweet potato. Both plants have similar growth forms and climatic requirements, and sweet potato is a high value crop, and thus we recommend planting sweet potato in areas infested by M. micrantha, perhaps as part of a rotation involving more vulnerable crops. Sweet potato could even be planted in habitats such as waste areas not currently cultivated in order to reduce M. micrantha populations. Our results showed that various components of reproduction for M. micrantha were significantly reduced by suppression of plant growth; the original data is available online [57]. Flowering phenology was impacted by sweet potato competition, and delayed flowering phenology, reduced duration of flowering and fruiting and increased duration of bud formation resulted in reduced pollinator visits and seed set for M. micrantha. Finally, high cover of sweet potato shading M. micrantha plants also reduced pollinator visits, seeds number, and seedling populations of M. micrantha. Thus the mechanism by which sweet potato reduced sexual reproduction in M. micrantha was essentially twofold: causing a delay in flowering phenology and reducing pollinator visits. In addition to utilizing sweet potato, research in this study and other recent studies revealed that control of M. micrantha ideally should take place during the seedling period when M. micrantha is most vulnerable and should be comprehensive for optimal results, employing both chemical and cultural control. Thus in the case of our study region in southern Asia, the most effective timing of control is in the peak of sexual seedling emergence in May-June. The potential for utilizing a crop like sweet potato to compete with an invasive plant may well apply to many other agronomic settings where other management techniques (e.g., chemical control, mechanical control or classical biological control) are unreliable or are associated with environmental concerns. The scenario of controlling M. micrantha by utilizing a crop with a similar growth form may provide a useful model for similar management strategies in other systems.

Authors' contributions

SCS and FDZ conceived and designed the experiments; SCS, GFX, GMJ, SFL, YXY, ADC, and FDZ performed the experiments; SCS and DRC analyzed the data and wrote the draft; HKN designed and commented on the manuscript. All authors read and approved the final manuscript.

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Competing interests

All authors declare that they have no competing interests.

Availability of supporting data

The data set supporting the results of this article is available in the Dryad Digital Repository http://dx.doi.org/10.5061/dryad.s2v53.

Ethics

All aspects of the study comply with institutional, national, and international guidelines. All experiments were conducted on non-regulated organisms. The study site was rented by Agricultural Environment and Resource Research Institute, Yunnan Academy of Agricultural Sciences and no permits were required to take samples.

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References

- Alpert P, Bone E, Holzapfel C. Invasiveness, invisibility and the role of environmental stress in the spread of nonnative plants. Perspect Plant Ecol Evol Syst. 2000;3:52–66. doi:10.1078/1433-8319-00004.
- Richardson DM, Allsopp N, D'Antonio CM, Milton SJ, Rejmánek M. Plant invasions—the role of mutualisms. Biol Rev. 2000;75:65–93. doi:10.1111/ i.1469-185X.1999.tb00041 x.
- Swamy PS, Ramakrishnan PS. Effect of fire on population dynamics of Mikania micrantha H.B.K. during early succession after slash-and-burn agriculture (Jhum) in northeast India. Weed Res. 1987;27:397–404. doi:10.1111/j.1365-3180.1987.tb01590.x.
- Britton AJ, Marrs RH, Carey PD, Pakeman RJ. Comparison of techniques to increase *Calluna vulgaris* cover on heath land invaded by grasses in Breckland, south east England. Biol Conserv. 2000;95:227–32. doi:10.1016/ S0006-3207(00)00047-1.
- Timmins SM. How weed lists help protect native biodiversity in New Zealand. Weed Technol. 2004;18:1292–300. doi:10.1614/0890-037X(2004) 018[1292:HWI HPNI2.0.CO:2.
- Paynter Q, Flanagan GJ. Integrating herbicide and mechanical control treatments with fire and biological control to manage an invasive wetland shrub, *Mimosa pigra*. J Appl Ecol. 2004;41:615–29. doi:10.1111/i.0021-8901.2004.00931.x.
- Shen SC, Xu GF, Zhang FD, Jin GM, Liu SF, Liu MY, Chen AD, Zhang YH. Harmful effects and chemical control study of *Mikania micrantha* H.B.K. in Yunnan, Southwest China. Afr J Agr Res. 2013;8:5554–61. doi:10.5897/ AJAR2013.7688.
- Barreto RW, Evans HC. The mycobiota of the weed Mikrania micrantha in southern Brazil with particular reference to fungal pathogens for biological control. Mycol Res. 1995;99:343–52. doi:10.1016/ S0953-7562(09)80911-8.

Shen et al. BMC Ecol (2016) 16:30 Page 8 of 9

- Mack RN, Simberloff D, Lonsdale WM, Evans H, Clout M, Bazzaz FA. Biotic invasions: causes, epidemiology, global consequences, and control. Ecol Appl. 2000;10:689–710. doi:10.1890/1051-0761(2000)010[0689:BICEGC]2. 0.CO:2.
- Messing RH, Wright MG. Biological control of invasive species: solution or pollution? Front Ecol Environ. 2006;4:132–40. doi:10.1890/1540-9295(2006)004[0132:BCOISS]2.0.CO:2.
- Yu H, Liu J, He WM, Miao SL, Dong M. Cuscuta australis restrains three exotic invasive plants and benefits native species. Biol Invasions. 2011;13:747–56. doi:10.1007/s10530-010-9865-x.
- Marrs RH, Frost AJ. A microcosm approach to the detection of the effects of herbicide spray drift in plant communities. J Environ Manag. 1997;50:369–88. doi:10.1006/jema.1996.9984.
- Milligan AL, Putwain PD, Marrs RH. A field assessment of the role of selective herbicides in the restoration of British moorland dominated by *Molinia*. Biol Conserv. 2003;109:369–79. doi:10.1016/S0006-3207(02)00163-5.
- Mason TJ, French K. Management regimes for a plant invader differentially impact resident communities. Biol Conserv. 2007;136:246–59. doi:10.1016/j.biocon.2006.11.023.
- 15. Ding H, Xu HG, Liu ZL. Impacts of invasion of *Eupatorium adenophorum* on vegetation diversity. J Ecol Rural Environ. 2007;23:29–32.
- Liu J, Wisniewski M, Droby S, Tian SP, Hershkovitz V, Tworkoski T. Effect of heat shock treatment on stress tolerance and biocontrol efficacy of Metschnikowia fructicola. Microbiol Ecol. 2011;76:145–55. doi:10.1111/j.1574-6941.2010.01037.x.
- Lugo AE. The apparent paradox of reestablishing species richness on degraded lands with tree monocultures. For Ecol Manag. 1997;99:9–19. doi:10.1016/S0378-1127(97)00191-6.
- Shen SC, Xu GF, Clements DR, Jin GM, Chen AD, Zhang FD, Hisashi KN. Suppression of the invasive plant mile-a-minute (*Mikania micrantha*) by local crop sweet potato (*Ipomoea batatas*) by means of higher growth rate and competition for soil nutrients. BMC Ecol. 2015;15:1–10. doi:10.1186/s12898-014-0033-5.
- Li WH, Luo JN, Tian XS, Chow WS, Sun ZY, Zhang TJ, Peng SL, Peng CL. A new strategy for controlling invasive weeds: selecting valuable native plants to defeat them. Sci Rep. 2015;5:11004. doi:10.1038/srep11004.
- Cao YS, Wang T, Xiao YA, Zhou B. The interspecific competition between Humulus scandens and Alternanthera philoxeroides. J Plant Interact. 2014;9:194–9. doi:10.1080/17429145.2013.808767.
- Gosper CR, Vivian-Smith G. Approaches to selecting native plant replacements for fleshy-fruited invasive species. Restor Ecol. 2009;17:196–204. doi:10.1111/j.1526-100X.2008.00374.x.
- Zhang LY, Ye WH, Cao HL, Feng HL. Mikania micrantha H.B.K. in China—an overview. Weed Res. 2004;44:42–9. doi:10.1111/j.1365-3180.2003.00371.x.
- Lowe S, Browne M, Boudjelas S, Poorter MD. 100 of the World's Worst Invasive Alien Species. A Selection from the Global Invasive Species Database. Auckland: IUCN/SSC Invasive Species Specialist Group (ISSG); 2001.
- Manrique V, Diaz R, Cuda JP, Overholt WA. Suitability of a new plant invader as a target for biological control in Florida. Invas Plant Sci Mana. 2011;4:1–10. doi:10.1614/IPSM-D-10-00040.1.
- Day MD, Kawi A, Kurika K, Dewhurst CF, Waisale S, Saul-Maora J, Fidelis J, Bokosou J, Moxon J, Orapa W, Senaratne KAD. *Mikania micrantha* Kunth (Asteraceae) (mile-a-minute): its distribution and physical and socioeconomic impacts in Papua New Guinea. Pac Sci. 2012;66:213–23. doi:10.2984/66.2.8.
- 26. Zan QJ, Wang YJ, Wang BS, Liao WB, Li MG. The distribution and harm of the exotic weed *Mikania micrantha*. Chin J Ecol. 2000;19:58–61.
- Shen SC, Xu GF, Clements DR, Jin GM, Liu SF, Zhang FD, Yang YX, Chen AD, Hisashi KN. Effects of invasive plant *Mikania micrantha* on plant community and diversity in farming systems. Asian J Plant Sci. 2015;14:27–33. doi:10.3923/ajps.2015.27.33.
- Yu H, Liu J, He WM, Miao SL, Dong M. Native Cuscuta campestris restrains exotic Mikania micrantha and enhances soil resources beneficial to natives in the invaded communities. Biol Invasions. 2009;11:835–44. doi:10.1007/s10530-008-9297-z.
- Xu GF, Zhang FD, Li TL, Shen SC, Zhang YH. Effects of 5 species and planting density on *Mikania micrantha* H.B.K growth and competitive traits. Ecol Environ Sci. 2011;20:798–804.
- 30. Shen SC, Xu GF, Zhang FD, Li TL, Zhang YH. Competitive effect of *Ipomoea batatas* to *Mikania micrantha*. Chin J Ecol. 2012;31:850–5.

- 31. Shen SC, Xu GF, Clements DR, Jin GM, Liu SF, Chen AD, Zhang FD, Hisashi KN. Control of invasive plant mile-a-minute (*Mikania micrantha*) with the local crop sweet potato (*Ipomoea batatas*) and applications of the herbicide bentazon. Asian J Plant Sci. 2014;13:59–65. doi:10.3923/ajps.2014.59.65.
- Kuorr YL, Chen TY, Lingg CC. Using a periodic cutting method and allelopathy to control the invasive vine, *Mikania micrantha* H.B.K. Taiwan J For Sci. 2002;17:171–81.
- Sihachakr D, Haïcour R, Cavalcante JM, Umboh I, Nzoghé D, Servaes A, Servaes A, Ducreux G. Plant regeneration in sweet potato (*lpomoea batatas* L., Convolvulaceae). Euphytica. 1997;96:143–52. doi:10.102 3/A:1002997319342
- De Wit CT. On Competition. Verslagen Landbouwkundige Onderzoekigen. 1960;66:1–82.
- Hoagland DR, Arnon DI. The water-culture method for growing plants without soil. Agricultural Experiment Station Circular. Berkley: College of Agriculture University of California; 1950.
- Hong L, Shen H, Ye WH, Cao HL. Study of pollinating insects of Mikania micrantha H.B.K and their foraging behavior. J South China Normal Univ (Nat Sci Ed). 2011;43:98–102. doi:10.6054/j. jscnun.2011.11.016.
- Liu ZH, Du GZ, Chen JK. Size-dependent reproductive allocation of *Ligularia virgaurea* in different habitats. Acta Phytoeol Sin. 2002:26:44–50.
- 38. Obeso JR. The cost of reproduction in plants. New Phytol. 2002;155:321–48. doi:10.1046/j.1469-8137.2002.00477.x.
- Lodge DM. Biological invasions: lessens for ecology. Trends Ecol Evol. 1993;8:133–7. doi:10.1016/0169-5347(93)90025-K.
- Van Kleunen M, Weber E, Fischer M. A meta-analysis of trait differences between invasive and non-invasive plant species. Ecol Lett. 2010;13:235– 45. doi:10.1111/j.1461-0248.2009.01418.x.
- 41. Reichard SH, Hamilton CW. Predicting invasions of woody plants introduced into North America. Conserv Biol. 1997;11:193–203. doi:10.1046/j.1523-1739.1997.95473.x.
- Wilke BJ, Irwin RE. Variation in the phenology and abundance of flowering by native and exotic plants in subalpine meadows. Biol Invasions. 2010;12:2363–72. doi:10.1007/s10530-009-9649-3.
- 43. Rejmánek M. A theory of seed plant invasiveness: the first sketch. Biol Conserv. 1996;78:171–81. doi:10.1016/0006-3207(96)00026-2.
- McIntosh ME. Flowering phenology and reproductive output in two sister species of *Ferocactus* (Cactaceae). Plant Ecol. 2002;159:1–13. doi:10. 1023/A:1015589002987.
- 45. Hong L, Shen H, Ye WH. Self-incompatibility in *Mikania micrantha* in South China. Weed Res. 2007;47:280–3. doi:10.1111/j.1365-3180.2007.00575.x.
- 46. Chittka L, Schürkens S. Successful invasion of a floral market. Nature. 2001;411:653. doi:10.1038/35079676.
- Bartomeus I, Vilà M, Santamaría L. Contrasting effects of invasive plants in plant-pollinator networks. Oecologia. 2008;155:761–70. doi:10.1007/ s00442-007-0946-1.
- Morales CL, Traveset A. A meta-analysis of impacts of alien vs. native plants on pollinator visitation and reproductive success of co-flowering native plants. Ecol Lett. 2009;12:716–28. doi:10.1111/j.1461-0248.2009.01319.x.
- Macanawai AR, Day MD, Tumaneng-Diete T, Adkins SW. The impact of rainfall upon pollination and reproduction of Mikania micrantha in Viti Levu, Fiji. Proceedings of the 23rd Asian-Pacific Weed Science Society Conference. Cairns; 2011.
- Ashman TL, Knight TM, Steets JA, Amarasekare P, Burd M, Campbell DR, Dudash MR, Johnston MO, Mazer SJ, Mitchell RJ, Morgan MT, Wilson WG. Pollen limitation of plant reproduction: ecological and evolutionary causes and consequences. Ecology. 2004;85:2408–21. doi:10.1890/03-8024.
- Yang QH, Feng HL, Ye WH, Cao HL, Deng X, Xu KY. An investigation of the effects of environmental factors on the flowering and seed setting of Mikania micrantha H.B.K (Compositae). J Trop Subtrop Bot. 2003;11:123–6.
- Zhang WY, Li GM, Wang BS, Zan QJ, Wang YJ. Seed production characteristics of an exotic weed *Mikania micrantha*. J Wuhan Bot Res. 2003;21:143–7.
- Hu YJ, But PPH. A study on life cycle and response to herbicides on Mikania micrantha. Acta Sci Nat Univ Sunvatseni. 1994:33:88–95.

Shen et al. BMC Ecol (2016) 16:30 Page 9 of 9

- 54. Zhang WY, Wang BS, Zhang JL, Li MG, Zan QJ, Wang YJ. Study on the structure and dynamics of seedlings of *Mikania micrantha* populations. Acta Sci Nat Univ Sunyatseni. 2002;41:64–6.
- 55. Shen SC, Xu GF, Zhang FD, Li TL, Jin GM, Zhang YH. Characteristics of the seed banks and seedling banks of *Mikania micrantha*-invaded soils different in type of habitat. J Ecol Rural Environ. 2013;29:483–8.
- Li X, Shen Y, Huang Q, Fan Z, Huang D. Regeneration capacity of small clonal fragments of the invasive *Mikania micrantha* H.B.K.: effects of burial depth and stolon internode length. PLoS ONE. 2013;8:e84657. doi:10.1371/journal.pone.0084657.
- Shen Sh-C, Xu G-F, Clements DR, Jin G-M, Yang Y, Zhang F-D, Kato-Nuguchi H. Data from: suppression of reproductive characteristics of the invasive plant Mikania micrantha by sweet potato competition. Dryad Digital Repository. 2016. doi:10.5061/dryad.9522r.

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